CDK4/6 inhibition protects normal cells against cancer therapy-induced damage

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Multi-agent platinum and radiation regimens, the preferred treatment for advanced malignancy (1), have increased cancer survivor rates, but the higher doses that may be required to stop the cancer can be toxic to normal tissues (2–6). This cancer therapy induced non-target tissue toxicity is a major limitation in cancer treatment and normal tissue toxicity the "*silent killer*" of many cancer survivors. The toxicity appears during or post cancer therapy and leads to effects that range from subclinical dysfunction to irreversible organ failure or even death due to impaired vital organ function and its metabolism (Figure 1). The issue is a growing concern as the number of cancer survivors is projected to increase from 16.9 million in 2019 to 26.1 million by 2040 (7).

Current technologies are not efficient enough to detect early diagnostic markers for cancer therapy induced normal tissue toxicity (8). Furthermore, no FDA-approved cancer drugs or interventional or combination therapies provide both normal tissue protection and inhibit cancer growth. It is therefore clinically warranted to employ high throughput technologies to unmask and identify the cancer therapy induced gene signatures responsible for early and late normal tissue toxicity and to develop a novel therapeutic strategy to simultaneously protect normal tissue from cancer therapy induced toxicities and control cancer growth.

Large data including bioinformatic on clinical samples and other research documents have demonstrated that how chemo-radiation induces mitochondrial reactive oxygen species (ROS) (9) and promotes normal tissue DNA damage through oxidative stress (6). This damage in turn leads to deregulation of the tumor suppressor retinoblastoma tumor suppressor RB/E2F pathway (10,11), which can result in DNA damage associated cancer

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progression, and ultimately to irreversible vital organ failure or death (12,13). Cancer discoveries suggest that cancer cell proliferation and migration is tightly regulated by the tumor suppressor retinoblastoma gene (RB1) and its pathway. Retinoblastoma protein belongs to the pocket protein family which includes p07, p110 and p130, and the deregulated retinoblastoma protein pathway is observed in many tumor models. RB controls the excessive proliferation via transcriptional repression of E2F target genes at the G1 to S phase transition. Under pathological conditions retinoblastoma protein pRb is inactivated (hyperphosphorylated) by cyclin dependent kinases (CDK4/6), which are serine/threonine kinases. Tumor suppressor pRb harbors multiple serine residues and CyclinD/CDK4/6 complex phosphorylates pRb at serine residues, and hyper-phosphorylated Rb becomes inactive and fails to induce transcriptional repression on cell cycle gene promoters. Pfizer Global Research made a long-standing effort to discover a CDK4/6 inhibitor called palbociclib (PD 0332991) that selectively inhibits CDK4/6 activity and in a joint effort with Fry *et al.* successfully formulated and tested it on RB proficient tumor models (14). The results from the study clearly demonstrated that palbociclib inhibited tumor growth in multiple tumor models in an RB proficient clinical setting.

Palbociclib functions as a potent antiproliferative agent in retinoblastoma protein (pRb)-positive tumor cells *in vitro* and *in vivo* and induces G1 arrest, with reduction in phospho-Ser780/Ser795 residues on pRb protein. Following the palbociclib discovery, additional CDK4/6 inhibitors were discovered: ribociclib, LEE 011, abemaciclib (LY2835219), and trilaciclib. Application of these CDK4/6 inhibitors on preclinical tumor models and use in clinical trials as single agent or fused with chemotherapy in patients with RB-positive tumors suggest that inhibition of CDK4/6 activity reestablishes cell cycle control by activating the pRb pathway. CDK4/6 inhibitors made to target ATP binding regions on CDK4/6 molecule. Palbociclib and ribociclib shows very high affinity to CDK4/6 protein and beyond cell cycle control and palbociclib also induces senescence and apoptosis via RB dependent mechanisms in RB positive cancer cells (15) as described in Figure 1.

The main purpose of this commentary on myelopreservation with CDK4/6 inhibitor trilaciclib by Weiss *et al.* (16) is to bring attention to the cancer research community that CDK4/6 inhibitors are not limited to suppression but also functions as novel protectors for normal cells against cancer therapy induced toxicity. Outcomes from multiples studies on CDK4/6 inhibitors also support the findings by Weiss *et al.* (16).

Mechanistic studies on an FDA-approved palbociclib (PD 0332991) anti-cancer drug have demonstrated in many tumor models that it targets the cell cycle and selectively inhibits cancer growth by activating the retinoblastoma tumor suppressor protein (inhibiting serine phosphorylation) and its signaling pathway (15,17,18). Recent discovery on palbociclib suggests that palbociclib also functions as a novel protector of normal tissue against therapy induced toxicity via RB-dependent mechanisms (15,19–21) and these studies also supports the work by Weiss *et al.* (16). CDK4/6 inhibition activates tumor suppressor protein pRB and the activated RB interacts with many of its interacting partners and performs multiple vital functions other than tumor suppression (Figure 2). In parallel, platinum-based cancer therapy induced damage on hematopoietic stem and progenitor cells (HSPC) causes multi-lineage myelosuppression. An intravenous application of trilaciclib (CDK4/6 inhibitor) (16)

preserves HSPC and immune system function against chemotherapy (myelopreservation), and this discovery along with published documents together strongly supports that palbociclib triggers antitumor immunity as described in (21). Recent findings suggest that CDK4/6 inhibition triggers apoptosis in non-small cell lung cancer (15) via activation of the pRB pathway and that RB is localized to the nucleus (22) and involved in the DNA repair pathway via nonhomologous recombination process. These discoveries suggest that CDK4/6 inhibitors protect normal tissue from cancer therapy induced toxicity (19). Additionally, recent discoveries strongly suggest that the CDK4/6 inhibitors play a major role in controlling pulmonary hypertension (23) via the RB dependent pathway.

Cyclin-dependent kinase CDK4/6 plays a vital role in mammalian cell cycle regulation and it drives progression of cells into S phase (DNA synthesis phase) of cell division. In tumors, CDK4/6 activity deregulates the $p16^{INK4a}$-Rb pathway that leads to uncontrolled cell division and cancer cell proliferation. Retinoblastoma tumor suppressor protein interacts with hundreds of molecules, involves in DNA repair pathway, and maintains genome integrity. Recently, reversible CDK4/6 inhibitors (palbociclib and trilaciclib) were employed to protect the immune system from chemotherapy induced toxicity. Weiss *et al.* (16) showed chemotherapy tolerance in lung cancer patients with myelopreservation benefits. Similarly, pulmonary arterial hypertension is mediated via proliferation of pulmonary arterial smooth muscle cells (PASMCs) with high CDK4/6 activity and poor prognosis. Selective inhibition of CDK4/6 via palbociclib inhibits PASMC proliferation via RB/E2F pathway (23).

Recent preclinical studies on CDK4/6 inhibitors suggest that CDK4/6 inhibitors play a vital role in normal cell protection other than tumor suppression. Additionally, CDK4/6 inhibition in human subjects by Weiss *et al.* reflects that CDK4/6 inhibitors have defensive potential to fight against cancer and therapy induced toxicity, and also triggers anti-tumor immunity in preclinical models (16,21,23–25). All of these studies suggest that CDK4/6 specific inhibition offers normal tissue protection and supports the Weiss *et al.* discovery with regard to myelopreservation in lung cancer patients (16). Taken together, these studies suggest that CDK4/6 inhibitors control cancer cell growth and simultaneously provide normal tissue protection against cancer therapy induced toxicities. Further studies are warranted to define the mechanistic pathways involved in normal tissue protection with CDK4/6 inhibition and to further interrogate whether this reversible CDK4/6 inhibition causes any long term side effects. Additionally, it is important to apply high throughput RNA and DNA sequencing technologies to investigate whether CDK4/6 inhibition promotes any drug resistance or any irreversible phenotypic or genotypic changes in normal tissue.

Acknowledgments

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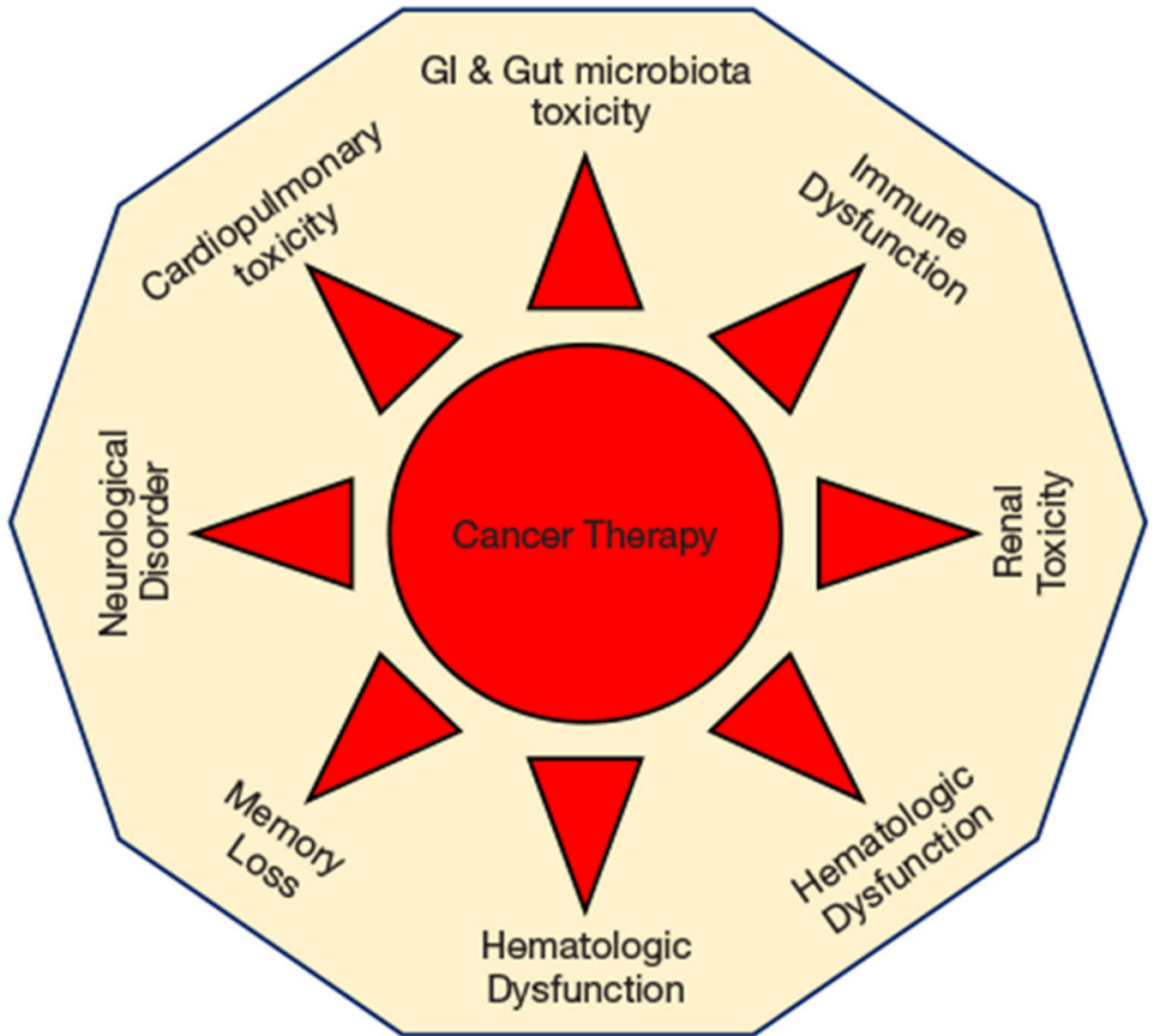
Figure 1.
Cancer therapy induced toxicity on vital organs. Diagrammatic representation of cancer therapy induced cardiopulmonary toxicity, neurological disorder, memory loss, hematological dysfunction, bone fracture, renal toxicity, and immune dysfunction, gastrointestinal intestinal and gut microbiota toxicity in cancer patients.
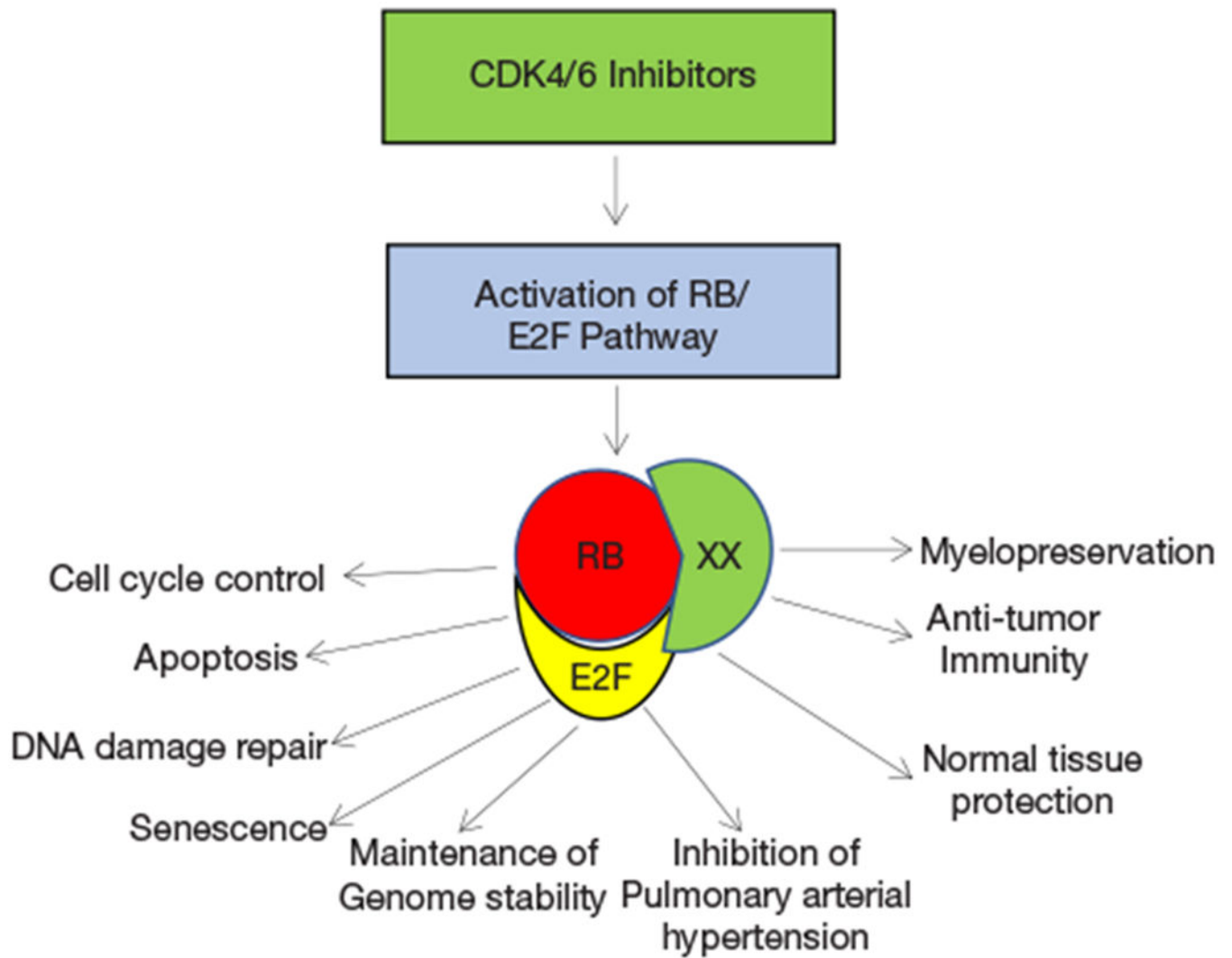
Figure 2.
Tumor suppressive and normal tissue protective potential of CDK4/6 inhibitors. CDK4/6 inhibitors activate RB/E2F signaling pathway and targets cell cycle machinery and induces apoptosis and cellular senescence. Additionally, CDK4/6 inhibitors involved maintaining genome stability by DNA repair mechanisms via RB dependent mechanism. CDK4/6 inhibition induces anti-tumor immunity and offers vital organ/cell protection, differentiation and myogenesis against cancer therapy induced toxicity.