

KINO 5 ANIMAL IMAGER

Basic Operation

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System Basics

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Start Up

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Software Overview

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EASY MODE

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BioLuminescence

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Fluorescence

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FAQ

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Sanitization



Shared Use means Shared Responsibility

REFILL empty or low Isoflurane

CLEAN all surfaces before and after use

SAVE files to external drive and remove from PC

System Basics

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Imaging Chamber

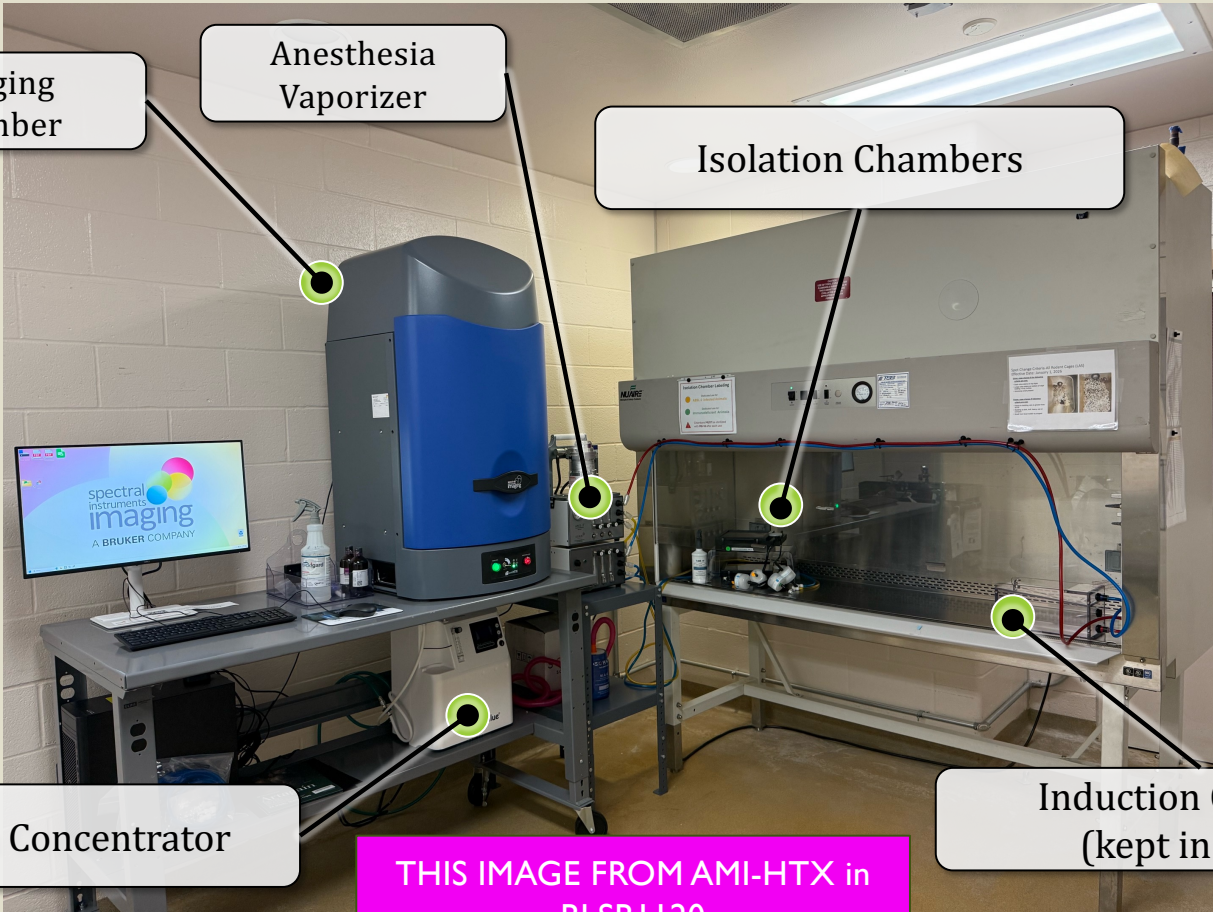
Anesthesia Vaporizer

Isolation Chambers

Oxygen Concentrator

Induction Chamber (kept in hood)

THIS IMAGE FROM AMI-HTX in BLSBI 120



Start Up

Manifold

Isoflurane

Oxygen

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- 1- Turn on SYSTEM
- 2- Each manifold port has separate flow control. Turn on and set the flow for the appropriate ports based on your imaging

- 3- Turn on EVAC timer
- 4- Set evac pump rate for each used port. EVAC rate should be roughly 10x input flow rate



- A) Induction
- B) AMI Manifold
- C) Isolation Chamber

All BLUE hoses are evac lines

Start Up

Manifold

Isoflurane

Oxygen

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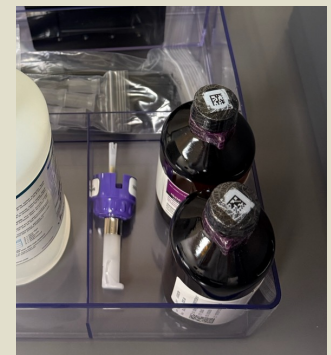
8



- 1- Ensure isoflurane level is between limits in clear window
- 2- Set dial to appropriate concentration



IF ISOFLURANE IS LOW – REFILL it using the provided stock bottles and attachable spout located next to the AMI.



Start Up

Manifold

Isoflurane

Oxygen



The AMI system uses an **oxygen concentrator** in place of consumable tanks.

Power on **AFTER** the flow rates are set on the manifold.

If no flow is on, the concentrator alarm will sound.

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Start Up

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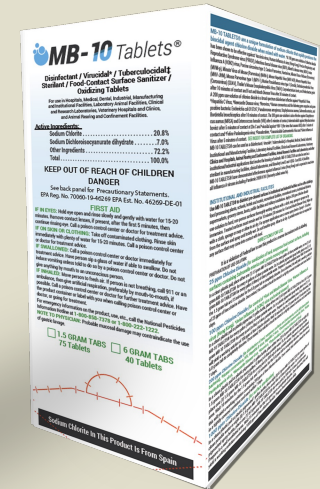
Animal Imagers and all associated equipment and surfaces must be sanitized **before** and **after** use with the provided PEROXIGARD. There are two (2) caveats to this:

1 – **DO NOT** spray Peroxigard directly into the AMI imaging chamber. Spray a paper towel and use that to disinfect all surfaces inside

2- **DO NOT** use Peroxigard on the isolation chambers as this may cloud the imaging window. PLEASE use the solution of MB-10 provided.



*Please use MB-10 test strips to confirm >100ppm before use



FOR ISOLATION CHAMBERS

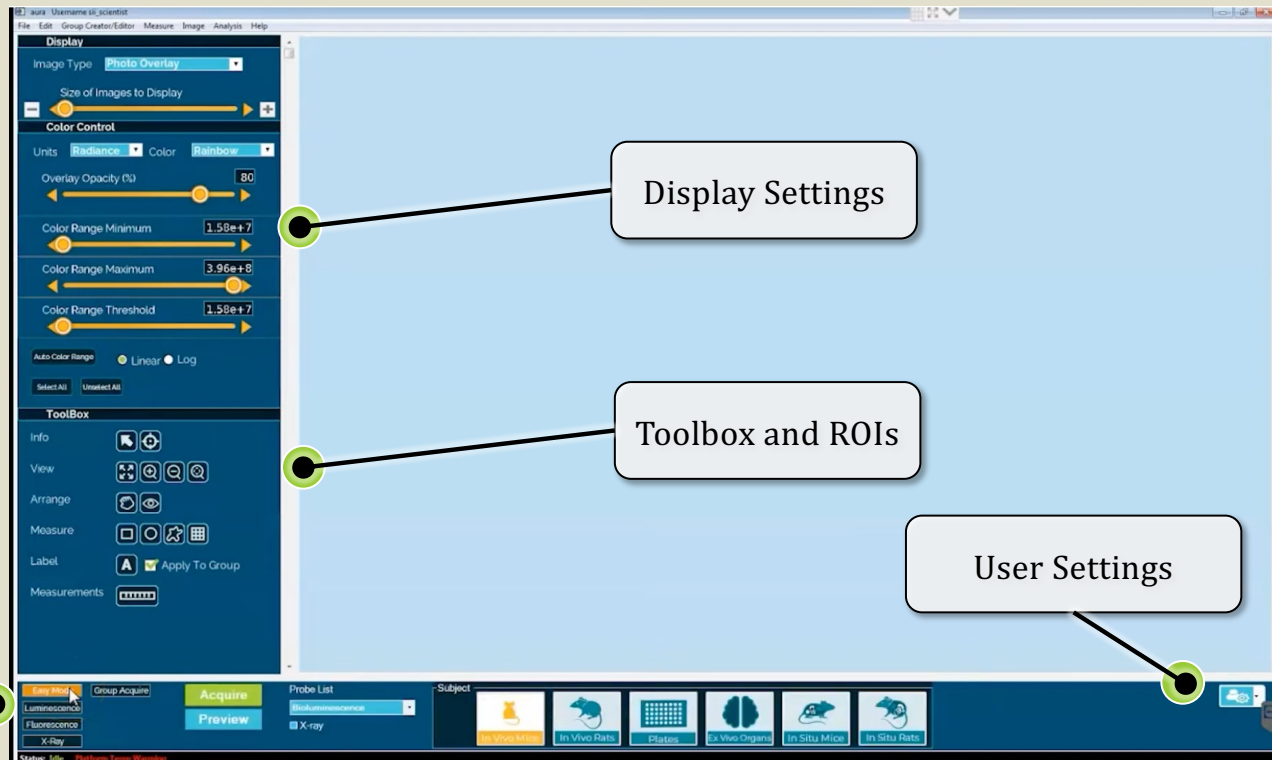
Software Overview

When logging into the PC
– Password is “**admin**”

When opening the software enter the
user name you created.

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Acquisition Modes
and Settings



Display Settings

Toolbox and ROIs

User Settings

EASY MODE

BioImaging Shared Resource - 2026

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The screenshot displays the EASY Mode software interface. On the left is a dark blue sidebar with various control panels: 'Display' (Image Type: Photo Overlay, Size of Images to Display slider), 'Color Control' (Units: Radiance, Color: Rainbow, Overlay Opacity: 80%, Color Range Minimum: 1.58e+7, Color Range Maximum: 3.96e+8, Color Range Threshold: 1.58e+7, Auto Color Range: Linear/Log), and 'ToolBox' (Info, View, Arrange, Measure, Label, Measurements). The main window is light blue and contains a large QR code. Below the QR code is the text 'YouTube: [EASY Mode tutorial](#)'. At the bottom, there is a navigation bar with 'Easy Mode' selected, 'Group Acquire', 'Acquire', 'Preview', 'Probe List' (Bioluminescence, X-ray), and 'Subject' (In Vivo Mice, In Vivo Rats, Plates, Ex Vivo Organs, In Situ Mice, In Situ Rats).

[YouTube: EASY Mode tutorial](#)

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The screenshot displays the auro software interface. On the left is a 'Display' panel with settings for 'Image Type' (Photo Overlay), 'Color Control' (Units: Radiance, Color: Rainbow, Overlay Opacity: 80%, Color Range Minimum: 1.54e+7, Color Range Maximum: 3.85e+8, Color Range Threshold: 1.54e+7), and 'ToolBox' (Info, View, Arrange, Measure, Label, Measurements). The main area contains two QR codes. Below the left QR code is the text 'YouTube: BLI Mode tutorial' and below the right QR code is 'YouTube: Kinetics through Group Acquire'. At the bottom, there is an 'Acquire' panel with settings for Exposure (1s), Binning (Low), FStop (1.2), Emission (Open), Field Of View (FOV: 20, 15, 10, 6), and Object Height (1.5 cm). A status bar at the very bottom indicates 'Status: Idle' and 'Fluorescence Temp Warning'.

Fluorescence

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The screenshot displays a software interface for fluorescence microscopy. On the left side, there is a vertical toolbar with icons for various functions: Info, View, Arrange, Measure, Label, and Measurements. The main window shows a large QR code centered on a light blue background. Below the QR code, the text "YouTube: FLI Mode tutorial" is displayed in green. The bottom of the interface features a control panel with various acquisition parameters such as Exposure (s), Binning, FStop, Excitation, Emission, Field Of View (FOV), and Object Height. The status bar at the very bottom indicates "Status: Idle" and "Platform Temp: Warning".

[YouTube: FLI Mode tutorial](#)

Troubleshooting

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1. The AMI unit has an **EMERGENCY STOP** button located on the front.
2. When pressed, the button disrupts all communication between the AMI and the PC
3. To **restore communication**, you must reset the button and re-initialize the software



THIS IMAGE FROM AMI-HTX in
BLSBI 120

SANITIZATION

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Animal Imagers and all associated equipment and surfaces must be sanitized **before** and **after** use with the provided PEROXIGARD. There are two (2) caveats to this:

2

1 – **DO NOT** spray Peroxigard directly into the AMI imaging chamber. Spray a paper towel and use that to disinfect all surfaces inside

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2- **DO NOT** use Peroxigard on the isolation chambers as this may cloud the imaging window. PLEASE use the solution of MB-10 provided.

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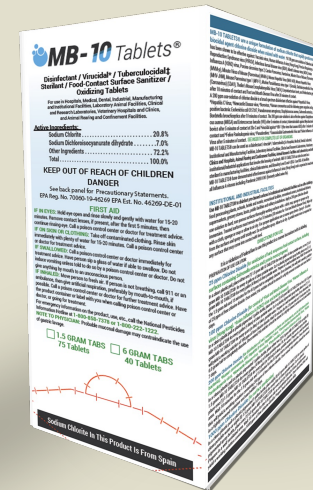
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FOR ISOLATION CHAMBERS